

Available online at www.sciencedirect.com



Tetrahedron Letters 45 (2004) 6895-6898

Tetrahedron Letters

New potent insecticidal agent: 4'-fucosyl avermectin derivative

Guohua Wei,^a Yuguo Du^{a,*} and Robert J. Linhardt^{b,*}

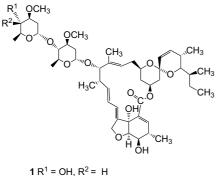
^aResearch Center for Eco-Environmental Sciences, Chinese Academy of Sciences, Beijing 100085, China ^bDepartments of Chemistry, Biology, and Chemical and Biological Engineering, Rensselaer Polytechnic Institute, Troy, NY 12180, USA

> Received 2 July 2004; revised 20 July 2004; accepted 21 July 2004 Available online 7 August 2004

Abstract—A 4'-fucosyl avermectin derivative was designed and synthesized. This new avermectin derivative showed excellent in vivo bioactivity against cabbage larvae when compared to commercially available avermectin B_{1a} . In this synthesis, thioglycosyl donors, but not trichloroacetimidates, were found compatible with sugar-macrolide synthesis under rt promotion with NIS or I₂ in *N*-meth-ylpyrrolidone.

© 2004 Published by Elsevier Ltd.

The avermectins are a family of naturally occurring macrocyclic lactones with exceedingly high activity against helminthes and arthropods.¹ A primary fermentation product of *Streptomyces avermitilis*, avermectin B_{1a} (1, AVM, Fig. 1), is an important and widely used agricultural pesticide.² Although compound 1 is extremely effective against mites, it is much less effective against insects, especially the cabbage looper, the core earworm and the southern armyworm.³ The level of activity against these species is insufficient to justify commercial development for these uses. Extensive inves-



 $2 R^{1} = H, R^{2} = MeNH \cdot HCO_{2}Ph$

Figure 1. Structures of AVM (1) and MK-244 (2).

* Corresponding authors. Tel.: +1-51827-63404; fax: +1-51827-63405; e-mail: linhar@rpi.edu

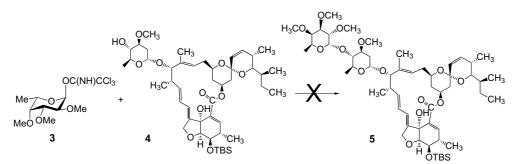
0040-4039/\$ - see front matter @ 2004 Published by Elsevier Ltd. doi:10.1016/j.tetlet.2004.07.092

tigation of the synthesis and biological evaluation of avermectin derivatives has been undertaken to obtain compounds with improved insecticidal activity.⁴ From these efforts, a major breakthrough came with the discovery of 4"-aminoavermectins.^{3a,5} These aminosugarcontaining avermectins showed excellent activity against a variety of insect larvae, spider mites and aphids. The use of 4"-epi-(methylamino)-4"-deoxyavermectin B_{1a} benzoate (**2**, MK-244, Fig. 1) as an agriculture insecticide has achieved commercial success.⁶ This specific example, when taken together with the success of other analogues,⁷ demonstrates that synthetic modifications at the terminal sugar of AVM offers derivatives having potent and improved bioactivity.

Our interest in these complex natural products led us to design a new AVM derivative in which 4'-hydroxyl of the oleandrosyl unit was replaced with a fully methylated α -L-fucopyranosyl moiety. L-Fucopyranosyl trichloroacetimidate donor **3** (Scheme 1) was selected for glycosylation of a modified avermectin lactone **4**⁸ under TMSOTf promotion in anhydrous CH₂Cl₂ to synthesize this target. To our surprise, sluggish glycosylation results were obtained throughout a wide range of solvents and reaction temperatures.⁹ Additional investigation showed that the macrolactone **4**, in the absence of donor, was highly unstable and decomposed quickly in the presence of catalytic amount of TMSOTf or 1 equiv of BF₃·Et₂O in CH₂Cl₂.

As most natural macrolide antibiotics contain sugar moieties,¹⁰ it was important to investigate the general coupling reaction conditions for sugar donor and

Keywords: Fucosyl avermectin; Avermectin; Insecticide; Synthesis; Glycosylation.



Scheme 1. Attempted synthesis of 5.

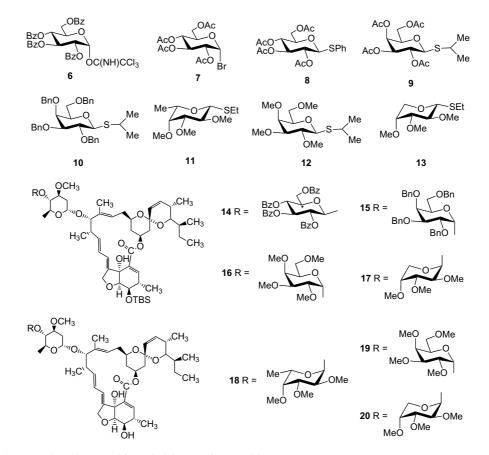


Figure 2. Glycosyl donors and products used in optimizing reaction conditions.

aglycone macrolactone. Thus, a number of glycosyl donors (Fig. 2) were prepared by routine methods and their reactive properties with lactone 4 were explored. AgOTf was a very compatible catalyst with trichloroacetimidate donor when compared to TMSOTf and BF₃. Et₂O.¹¹ AgOTf catalyzed glycosylation of glucopyranosyl trichloroacetimidate 6 and aglycon 4 in CH_2Cl_2 gave a very clean reaction with 91% yield of desired compound (see Table 1, entry 2). When the same reaction conditions were applied to the coupling of 3 and 4, a 50% yield of expected 5 was isolated as an α,β mixture with a ratio of 1:1 (Table 1, entries 3 and 4). We speculated that the rapid consumption of **3** was a critical factor in this low yielding reaction. Unfortunately, changing solvent from CH₂Cl₂ to toluene and performing the reaction at the reduced temperature did not improve the yield. AgOTf, in the presence or absence of lutidine, failed to catalyze the glycosylation of lactone **4** with bromide donor **7** (entries 5 and 6).

Thioglycosides have often been applied in macrolide antibiotics synthesis.¹² However, when thioglycosides **8** and **9** were used under promotion with MeOTf, NBS or NIS catalysts, no desired products were obtained (entries 7–10). These results suggested that fully acetyl protected donor might be deactivated. When a benzylated thioglycoside donor **10** was used in the presence of NIS, a 60% yield of product was isolated as an α : β mixture (entry 11). Encouraged by these results, together with the observation that most sugar residues found in antibiotics are methylated and deoxygenated,¹⁰ we prepared fucosyl thioglycoside donor **11** for the glycosyl-

Table 1. Glycosylation of sugar donor (3, 6–13) and macrol

Entry	Donor	Catalyst	Solvent	Result (isolated yield)
1	3	TMSOTf	CH ₂ Cl ₂	No desired product
2	6	AgOTf	CH ₂ Cl ₂	14 , 91%, β only
3	3	AgOTf	CH ₂ Cl ₂	5 , 50%, α : β = 1:1
4	3	AgOTf	Toluene	5 , 50%, $\alpha:\beta = 1:1$
5	7	AgOTf	CH_2Cl_2	No desired product
6	7	AgOTf/lutidine	CH_2Cl_2	No desired product
7	8	MeOTf	CH ₂ Cl ₂	No desired product
8	8	NBS	CH_2Cl_2	No desired product
9	9	NBS	CH_2Cl_2	No desired product
10	9	NIS	N-Methylpyrrolidone	No desired product
11	10	NIS	CH_2Cl_2	15 , 60%, $\alpha:\beta = 1:1$
12	11	NIS	CH_2Cl_2	5 , 45%, α : β = 1:1
13	10	NIS	N-Methylpyrrolidone	15 , 70%, α : β = 2:1
14	11	NIS	N-Methylpyrrolidone	5 , 86%, α : β = 4:1
15	11	I_2	N-Methylpyrrolidone	5 , 82%, α : β = 4:1
16	12	I_2	N-Methylpyrrolidone	16 , 80%, α : β = 7:3
17	13	I_2	N-Methylpyrrolidone	17 , 85%, $\alpha:\beta = 3:1$

ation of **4** under the same reaction conditions, and a 45% yield of desired product was obtained as a 1:1 α : β mixture (entry 12). Further investigation showed that both **10** and **11** could afford improved yields and stereo-selectivities in *N*-methylpyrrolidone (entries 13 and 14). A typical procedure is the following: To a solution of **11** (1 mmol) and **4** (0.9 mmol) in *N*-methylpyrrolidone (5 mL) was added NIS (1.1 equiv) under a N₂ atmosphere. The mixture was stirred at 25 °C for 1 h, then poured into water and extracted with CH₂Cl₂. A routine column separation gave α -linked **5** (86% total yield, α , β = 4:1) as an amorphous solid.

We next examined I₂ as a less expensive replacement for NIS, to lower the cost of making this potential pesticide, and found I₂ to be a suitable promoter for this reaction.¹³ When methylated glycosyl donors **11**, **12** and **13** were coupled with **4** in *N*-methylpyrrolidone under I₂ promotion, compounds **5**, **16** and **17** were afforded predominantly as the α -isomers in 82%, 80% and 85% yield, respectively.¹⁴ Finally, clean removal of the *tert*-butyldimethylsilyl (TBS) group from **5**, **16** and **17**, using hydrogen fluoride–pyridine complex, afforded the corresponding **18**, **19** and **20**, respectively. It is noteworthy that all attempts to cleave TBS with *tetra-n*-butylammonium fluoride, using published methods, failed.¹⁵

Bioactivity was evaluated in preliminary studies using the cabbage leaf dip bioassay described by Zhao et al.¹⁶ The fourth instar larvae were tested with compounds **18**, **19** and **20** in acetone solution and mortality was assessed 3 days after treatment. The results showed potency at microgram levels as summarized in Table 2. Toxicity in animals was examined in ICR mice using standard methods and the LD_{50} of compound **18** was 87.5 mg/kg.

In conclusion, a novel AVM analogue has been designed and synthesized. The key step is the stereoselective high yielding glycosylation of 4'-position of AVM derivative. The use of fully methylated thioglycosides as donors, and NIS or I_2 as catalyst in *N*-methylpyrrolidone at rt,

Table 2. Bioactivities of compounds 18, 19, 20 to cabbage larvae

Dosage	Microgram per larva	Total number	Dead number	Mortality (%)
18	2.0	90	84	93.3
19	2.0	90	77	85.5
20	2.0	90	68	75.5
1	2.0	90	48	53.3
2	2.0	90	83	92.2
Acetone	_	30	2	6.7

provided good yields of target AVM analogues, which show excellent bioactivity against cabbage larvae. The method described here should be valuable in the synthesis of other sugar-containing macrolide antibiotics.¹⁰

Acknowledgements

This work was supported by National Basic Research Program of China (2003CB415001), NNSF of China (20372081, 30330690), and NIH of the US (HL62244).

References and notes

- (a) Fisher, M.; Mrozik, H. In *Macrolide Antibiotics*; Omura, S., Ed.; Academic: New York, 1984, pp 553–606; (b) Davis, H. G.; Green, R. H. *Nat. Prod. Rep.*, **1986**, 87–121.
- (a) Dybas, R. A. In *Ivermectin and Abamectin*; Campbell, W. C., Ed.; Springer: New York, 1989, pp 287–310; (b) Campbell, W. C.; Fisher, M. H.; Stapley, E. O.; Albers-Schönberg, G.; Jacob, T. A. *Science* 1983, 221, 823–828.
- (a) Mrozik, H.; Eskola, P.; Linn, B. O.; Lusi, A.; Shih, T. L.; Tischler, M.; Waksmunski, F. S.; Wyvratt, M. J.; Hilton, N. J.; Anderson, T. E.; Babu, J. R.; Dybas, R. A.; Preiser, F. A.; Fisher, M. H. *Experientia* 1989, 45, 315–316; (b) Putter, I.; MacConnell, J. G.; Preiser, F. A.; Haidri, A. A.; Ristich, S. S.; Dybas, R. A. *Experientia* 1981, 37, 963–964.
- 4. (a) For leading references to synthetic studies see: Blizzard, T. A. Org. Prep. Proc. Int. 1994, 26, 645–670;
 (b) Davies, H. G.; Green, R. H. Chem. Soc. Rev. 1991, 20, 211–269; (c) Davies, H. G.; Green, R. H. Chem. Soc. Rev.

1991, 20, 271–339; (d) Hanessian, S.; Ugolini, A.; Hodges, P. J.; Beaulieu, P.; Dubé, D.; André, C. *Pure Appl. Chem.* **1987**, 59, 299–304; (e) White, J. D.; Bolton, G. L. *J. Am. Chem. Soc.* **1990**, *112*, 1626–1628; (f) White, J. D.; Bolton, G. L.; Dantanarayana, A. P.; Fox, C. M. J.; Hiner, R. N.; Jackson, R. W.; Sakuma, K.; Warrier, U. S. *J. Am. Chem. Soc.* **1995**, *117*, 1908–1939, and references cited therein.

- Dybas, R. A.; Hilton, N. J.; Babu, J. R.; Preiser, F. A.; Dolce, G. J. *Top. Ind. Microbiol.* **1989**, 203–212.
- (a) Fisher, M. H. Pure Appl. Chem. 1990, 62, 1231–1240;
 (b) Cvetovich, R. J.; Kelly, D. H.; DiMichele, L. M.; Shuman, R. F.; Grabowski, E. J. J. J. Org. Chem. 1994, 59, 7704–7708.
- (a) Meinke, P. T.; O'Connor, S. P.; Ostlind, D. A.; Shoop, W. L.; Fisher, M. H.; Mrozik, H. *Bioorg. Med. Chem. Lett.* 1993, *3*, 2675–2680; (b) Rohrer, S. P.; Meinke, P. T.; Hayes, E. C.; Mrozik, H. *Proc. Natl. Acad. Sci. U.S.A.* 1992, *89*, 4168–4172.
- 8. Compound 4 was obtained from the avermectin B_{1a} hydrolysis (1% H_2SO_4 in isopropyl alcohol), followed by selective silylation (TBSCl, Im, DMF).
- We explored CH₂Cl₂, ether, toluene, acetonitrile, nitromethane, hexane and THF as glycosylation solvents at temperatures ranging from -42°C to rt.
- (a) Steinmetz, W. E.; Shapiro, B. L.; Robert, J. J. J. Med. Chem. 2002, 45, 4899–4902; (b) Tanaka, T.; Yuji, O.; Hamada, T.; Yonemitsu, O. Tetrahedron Lett. 1986, 27, 3651–3654; (c) Peterson, I.; Mansuri, M. M. Tetrahedron 1985, 41, 3569–3624.
- 11. Wei, G.; Gu, G.; Du, Y. J. Carbohydr. Chem. 2003, 22, 385–393.
- (a) Nicolaou, K. C.; Seitz, S. P.; Papahatjis, D. P. J. Am. Chem. Soc. 1983, 105, 2430–2434; (b) White, J. D.; Blakemore, P. R.; Browder, C. C.; Hong, J.; Lincoln, C. M.; Nagornyy, P. A.; Robarge, L. A.; Wardrop, D. J. J. Am. Chem. Soc. 2001, 123, 8593–8595; (c) Loewe, M. F.; Cvetovich, R. J.; DiMichele, L. M.; Shuman, R. F.; Grabowski, E. J. J. J. Org. Chem. 1994, 59, 7870–7875; (d) Blizzard, T. A.; Margiatto, G. M.; Mrozik, H.; Shoop, W. L.; Frankshun, R. A.; Fisher, M. H. J. Med. Chem. 1992, 35, 3873–3878.

- I₂ has been used to promote glycosylation see: Kartha, K. P. R.; Aloui, M.; Field, R. A. *Tetrahedron Lett.* **1996**, *37*, 5175–5178.
- Physical data for compound 17: [α]_D²⁵ -52 (*c* 1, CHCl₃); ¹H NMR: (400 MHz, CDCl₃): 0.85–0.95 (m, 10H, CH₃-28, CH₃-26a, CH₃-14a, H-18a), 1.14 (d, 3H, CH₃-12a), 1.24-1.26 (m, 6H, CH₃-6^I, CH₃-6^{II}), 1.45–1.50 (m, 4H, CH₃-14a, H-20a), 1.58–1.64 (m, 4H, H-16a, H-2¹a, CH₂-27), 1.76 (m, 1H, H-18e), 1.88 (br s, 3H, CH₃-4a), 2.00-2.05 (m, 1H, H-20e), 2.23–2.29 (m, 4H, H-2¹e, H-24, H-26, H-16e), 2.35-2.37 (d, 1H, 5-OH), 2.47-2.53 (m, 1H, H-12), 3.28-3.30 (m, 1H, H-2), 3.35-3.41 (m, 1H, H-4^I), 3.42 (s, 31, 0CH₃), 3.46–3.50 (m, 2H, H-3^{II}, H-25), 3.53–3.60 (3 s, 9H, 0CH₃), 3.62–3.66 (dd, 1H, $J_{1,2}^{II,II}$ 4.1, $J_{3,2}^{II,II}$ 10.1 Hz, H-2^{II}), 3.78–4.00 (m, 7H, H-3^I, H-17, H-13, H-5^I, H-4^{II}, H-6, 7-OH), 4.20-4.23 (m, 1H, H-5^{II}), 4.28-4.32 (t, 1H, H-5), 4.68-4.74 (m, 3H, CH₂-8a, H-1^I), 4.96 (m, 1H, H-3), 5.37-5.43 (m, 2H, H-19, H-15), 5.54–5.57 (dd, 1H, $J_{23,22}$ 2.5, $J_{23,24}$ 9.9 Hz, H-23), 5.60 (d, 1H, $J_{1,2}^{II,II}$ 4.0 Hz, H-1^{II}), 5.70– 5.72 (t, 2H, H-10, H-11), 5.75–5.78 (dd, 1H, H-22), 5.85– 5.88 (m, 1H, H-9); ¹³C NMR (100 MHz, CDCl₃): 12.02 (C-28), 12.96 (C-26a), 15.07 (C-14a), 16.38 (C-24a), 16.47 (C-6^{II}), 18.50 (C-6^I), 19.97 (C-4a), 20.00 (C-12a), 27.50 (C-27), 30.58 (C-24), 34.26 (C-16), 34.50 (C-26), 35.17 (C-2^I), 36.60 (C-18), 39.83 (C-12), 40.49 (C-20), 45.72 (C-2), 56.07, 57.99, 58.87, 61.70, 66.51 (C-5^I), 66.97 (C-5^{II}), 66.74 (C-5), 68.15 (C-19), 68.33 (C-8a), 68.48 (C-17), 74.89 (C-25), 77.62 (C-2^{II}), 79.02 (C-3^I), 79.05 (C-4^{II}), 79.25 (C-4^I), 79.26 (C-6), 80.11 (C-3^{II}), 80.41 (C-7), 82.57 (C-13), 95.31 (C-1^I), 95.77 (C-21), 96.96 (C-1^{II}), 118.06 (C-15), 118.37 (C-3), 120.43 (C-9), 124.76 (C-10), 127.78 (C-23), 135.31 (C-14), 136.30 (C-22), 137.98 (C-4), 138.04 (C-11), 139.59(C-8), 173.72 (C-1); MALDITOF-MS: calcd for C₅₀H₇₆O₁₅, 916.5 [M]; found: 939.70 $[M+Na]^+$; 955.70 $[M+K]^+$.
- (a) Hanessian, S.; Ugolini, A.; Dubé, D.; André, C. J. Am. Chem. Soc. 1986, 108, 2776–2778; (b) Nicolaou, K. C.; Dolle, R. E.; Papahatjis, D. P.; Randall, J. L. J. Am. Chem. Soc. 1984, 106, 4189–4192.
- Zhao, J.-Z.; Bishop, B. A.; Grafius, E. J. J. Econ. Entomol. 2000, 93, 1508–1514.